

### Review Article

## A NOVEL TARGETED FORMULATION FOR OSTEOARTHRITIS: EXPLORING SYNERGISTIC BENEFITS OF *CISSUS* *QUADRANGULARIS*, *BOSWELLIA SERRATA*, PROPOLIS AND PALMITOYLETHANOLAMIDE

B.Dharani\*<sup>1</sup> , Suba.A<sup>1</sup> 

<sup>1</sup> Department of Physiology, Faculty of Medicine, A.C.S Medical College and Hospital, Dr.M.G.R Educational and Research Institute, Chennai, India.

\* Correspondence, e-mail: doctordharanibhaskaran@gmail.com

Received: 26.04.2025 / Revised: 14.07.2025 / Accepted: 19.08.2025 / Published online: 01.10.2025 /  
Published in final version: 14.01.2026

### ABSTRACT

Osteoarthritis (OA) is known as a debilitating form of arthritis that is marked by progressive degradation of cartilage, synovial inflammation, chronic pain, and subchondral bone remodeling. OA causes progressive stiffness and decreased mobility, significantly affecting the overall quality of life of the person affected. In spite of vast research in this area, the present pharmacological interventions are purely symptomatic. Consequently, there is an expanding interest in exploring multidimensional targeting of pathophysiological pathways using natural treatment options, while improving patient compliance by enhancing the safety profile. The current review focuses on a novel, innovative, and conceptual formulation that is designed by the authors with scientific-evidence-packed natural compounds for management of OA. This review aims to evaluate the rationale behind formulating a conceptual novel tablet consisting of *Cissus quadrangularis*, *Boswellia serrata*, propolis, and palmitoylethanolamide (PEA) for definitive management of OA. To our knowledge, this is the first article to explore this combination which is designed in such a way that it targets oxidative stress, inflammation, cartilage destruction, and pain in OA simultaneously in a synergistic manner. In contrast to conventional treatment options, which primarily provide symptom relief, this novel conceptual formulation could offer analgesic, chondroprotective, and regenerative effects with a reasonable safety profile, making it suitable for long-term use. This formulation has the potential to emerge as an effective and safer alternative for treatment of OA, by helping to bridge the gap between integrative and conventional medicine.

**KEYWORDS:** *Boswellia*, *Cissus*, Drug Formulation, Osteoarthritis, *Palmitoylethanolamide*, *propolis*

Article is published under the CC BY license.

### 1. Introduction

Osteoarthritis (OA) is a debilitating condition that is said to affect around 303 million people all over the world [1]. It consists of gradual cartilage degradation, matrix degeneration, inflammation of the synovium, subchondral remodeling of bone, and chronic intolerable pain. It leads to stiffness of the joint involved, associated with reduced joint mobility that causes a reduction in patient's quality of life [2]. Because of the rise in the aging population, obesity, and sedentary lifestyle, the prevalence of OA is predicted to increase. The currently available treatment options include drugs like Nonsteroidal Anti-Inflammatory Drugs (NSAIDs), corticosteroids, and major surgical

procedures like total joint replacement. However, these treatment options are purely symptomatic, offering temporary pain relief rather than providing a definitive cure [3,4]. Additionally, they present a risk of long-term complications. Recently, various studies are being conducted on Disease-Modifying Osteoarthritis Drugs (DMOADs) that have the potential to provide a curative treatment [5–7]. Consequently, there is an expanding interest in exploring multidimensional targeting of pathophysiological pathways using natural treatment options, while improving patient compliance by enhancing the safety profile. In recent years, numerous bioactive compounds of natural sources have shown significant therapeutic potential for OA [8]. Notably, *Cissus quadrangularis*, *Boswellia serrata*, propolis, and

palmitoylethanolamide (PEA) have emerged as promising treatment options for OA because of their potential antioxidant, anti-inflammatory, analgesic, and chondroprotective properties. *C. quadrangularis* is a traditionally used ancient treatment for joint and bone health that has been extensively researched for its positive role in decreasing joint inflammation and improving collagen synthesis [9,10]. *B. serrata* is another notable herbal extract that has rich anti-inflammatory activity, primarily through 5-lipoxygenase (5-LOX) inhibition which is an important enzyme in leukotriene synthesis [11]. Propolis is a bee product that is rich in polyphenols, possessing strong immunomodulatory and antioxidant properties [12]. PEA is an endogenous fatty acid amide that exerts potential anti-inflammatory and analgesic effects by activation of Peroxisome Proliferator-Activated Receptor- $\alpha$  (PPAR- $\alpha$ ) and regulation of mast cells [13,14]. Due to the intricate and multifaceted nature of OA, a combination strategy harnessing these bioactive compounds could offer a better treatment outcome compared to individual therapy. The current review focuses on a novel, innovative, and conceptual formulation that is designed by the authors with the scientific-evidence-packed natural compounds *C. quadrangularis*, *B. serrata*, propolis, and PEA for management of OA. To our knowledge, this is the first article to explore this combination. This review aims to evaluate the rationale behind formulating a novel conceptual tablet consisting of *C. quadrangularis*, *B. serrata*, propolis, and PEA for definitive management of OA, which might have a strong clinical translational potential. By assessing their pharmacological activities, possible synergistic effects, and mechanisms of action, we try to provide an evidence-based foundation for this novel formulation for OA management.

## 2. Methodology

This review was carried out to create and evaluate the science behind a conceptual evidence-based formulation targeting the pathophysiology of OA, consisting of *C. quadrangularis*, *B. serrata*, propolis, and PEA. A complete search of literature was done using the relevant keywords, and all related studies published during the period 2000–2024 exploring the mechanisms of action, pharmacological effects, safety profile, and synergistic interactions of these components were included, which was conducted using scientific repositories like SCOPUS, Google Scholar, PubMed, and Web of Science. Those articles published in other languages besides English, not focusing on OA, and duplicates were excluded. A summary of data was synthesized to derive the potential of these components in specifically targeting different pathophysiological pathways of OA like oxidative stress, inflammation, cartilage degradation, and pain. Based on the cumulative evidence, the current novel formulation was conceptualized to offer multifaceted therapeutic benefits comprising of anti-inflammatory, antioxidant, analgesic, and chondroprotective effects with a better safety profile in the context of long-term use. Even though this review was not written using software for systematic review, we have made efforts to reduce selection bias through a structured inclusion strategy and search. In further studies, software tools like Rayyan might be utilized to improve methodological rigor.

## 3. Pathophysiology of OA

Osteoarthritis is defined as a chronic form of degenerative joint disease that specifically affects the synovium, articular cartilage, and subchondral bone, eventually causing stiffness, pain, and impaired mobility of the joint. It was previously considered as a disease of mechanical wear and tear. However, evolving evidence indicates that it is primarily caused by an intricate biochemical and molecular pathway encompassing oxidative stress, inflammation, synovial destruction, cartilage degradation, and maladaptive pain signalling [15–17]. Interpretation of this complex pathological process is crucial in identifying the potential therapeutic targets, in order to provide a definitive treatment for the disease.

### 3.1. Inflammatory cascades

Inflammation has a critical role in the pathophysiology of OA. Inflammatory mediators like chemokines and cytokines destabilize the balance between catabolic (cartilage-degrading) and anabolic (cartilage-building) processes. This leads to formation of catabolic enzymes that are involved in the destruction of the joint [18,19]. The key cytokines that mediate the pathophysiology of OA are Tumor Necrosis Factor- $\alpha$  (TNF- $\alpha$ ) and Interleukin-1 $\beta$  (IL-1 $\beta$ ), which cause formation of proteases such as Matrix Metalloproteinases and aggrecanases [20–23]. Consequently, this leads to degradation of the Extracellular Matrix (ECM) and degradation of cartilage. Additionally, the key enzymes were found to mediate formation of cyclooxygenase (COX-2) and prostaglandins (PGE2) which further heightens the inflammation and pain [24, 25]. Hence, these cytokines are known as pro-inflammatory cytokines that cause the pathological changes in the joint [26]. Also, another critical mediator of inflammation was found to be the NLRP3 inflammasome, which links this process of inflammation with mechanical stress [27].

### 3.2. Oxidative stress

In a joint affected by OA, there exists an imbalance between antioxidant defense and oxidative stress [28, 29]. Because of inflammation, excess production of Reactive Oxygen Species (ROS) occurs in the joint [30]. This activates Nuclear Factor-kappa B (NF- $\kappa$ B) which is an essential transcription factor that increases pro-inflammatory cytokines, further intensifying the joint inflammation. Also, this induces MMPs that further degrade the Extracellular Matrix (ECM). Overall, the prevailing oxidative damage to synovial cells and chondrocytes causes apoptosis and mitochondrial dysfunction, which further increases cartilage breakdown [31].

### 3.3. Cartilage degradation

The existing imbalance between catabolic and anabolic mechanisms causes cartilage degradation, which is known as the hallmark of OA [32, 33]. Chondrocytes are the crucial cells in regulating ECM homeostasis [34]. Increased catabolic processes are marked by excess production of catabolic enzymes like MMP-1, MMP-3, and MMP-13 and a disintegrin and metalloproteinase with thrombospondin motifs-4 & -5 (ADAMTS-4 & -5), which degrade essential components of the ECM like aggrecan and type II collagen [35]. The loss of proteoglycans secondary to destruction of cartilage matrix reduces

cartilage hydration and disrupts its shock-absorbing property, making the joint vulnerable to further mechanical degradation [36, 37].

### 3.4. Synovial Inflammation

The inflammation of the synovium, known as synovitis, occurs in OA due to inflammation-mediated infiltration of immune cells like T cells and macrophages and elevated synovial fluid cytokines. Due to this condition, the inflamed synovial tissue produces inflammatory mediators like IL-6, TNF- $\alpha$ , and PGE2, which leads to synovial vascularization and hyperplasia [38–40]. This, in turn, leads to excess joint effusion, joint stiffness, and further accelerated cartilage degradation.

### 3.5. Pain mechanisms

The pain in OA involves multiple pathomechanisms, which include peripheral sensitization, central sensitization, neuropeptides, and subchondral bone remodelling. In OA, the inflammatory mediators such as prostaglandins, bradykinin, and cytokines sensitize the pain receptors in the joint, leading to reduced pain threshold, which is called peripheral sensitization [41, 42]. Apart from these, chronic pain causes specific neuroplastic changes that result in exacerbated pain stimuli [43]. Additionally, certain neuropeptides such as Calcitonin Gene-Related Peptide (CGRP) and Substance P that are released from nerve endings cause increased response to pain and neurogenic inflammation [44]. As a result of chronic inflammation, remodeling of subchondral bone occurs, leading to appearance of osteophytes, which further aggravates the nociceptive signalling process [45–47].

## 4. Proposed drug combination

In the current review, the proposed combination (Fig. 1) of drugs for the rationally designed tablet comprises *C. quadrangularis*, *B. serrata*, propolis and PEA. It is formulated in such a way that it could target various pathways of OA pathophysiology such as inflammation, oxidative stress, degradation of cartilage, and chronic pain in a synergistic way. This can offer a safer and more effective option for treatment of OA, by helping to bridge the treatment gap that exists between conventional medicine and integrative medicine. Table 1 shows the evidence-based critical analysis of the mechanism of formulation.

### 4.1. *C. quadrangularis*

It is a commonly used Ayurveda medicine for management of fractures, osteoporosis, and joint disorders. It is also called the “Bone Setter’s” plant that was found to have anti-inflammatory, antioxidant, regenerative, and chondroprotective actions, through regulation of pro-inflammatory cytokines, suppressing NF- $\kappa$ B pathway and activating alkaline phosphatase [48–50]. Previous studies have shown that it can significantly reduce joint pain [51]. Additionally, it activates synthesis of the matrix and proliferation of chondrocyte, thereby causing chondroprotective effects [52]. Further, it was found to decrease the release of pro-inflammatory cytokines and improve genes controlling differentiation of osteoblast [53]. These findings support the fact that it has a potential to protect cartilage and suppress inflammation in management of OA.

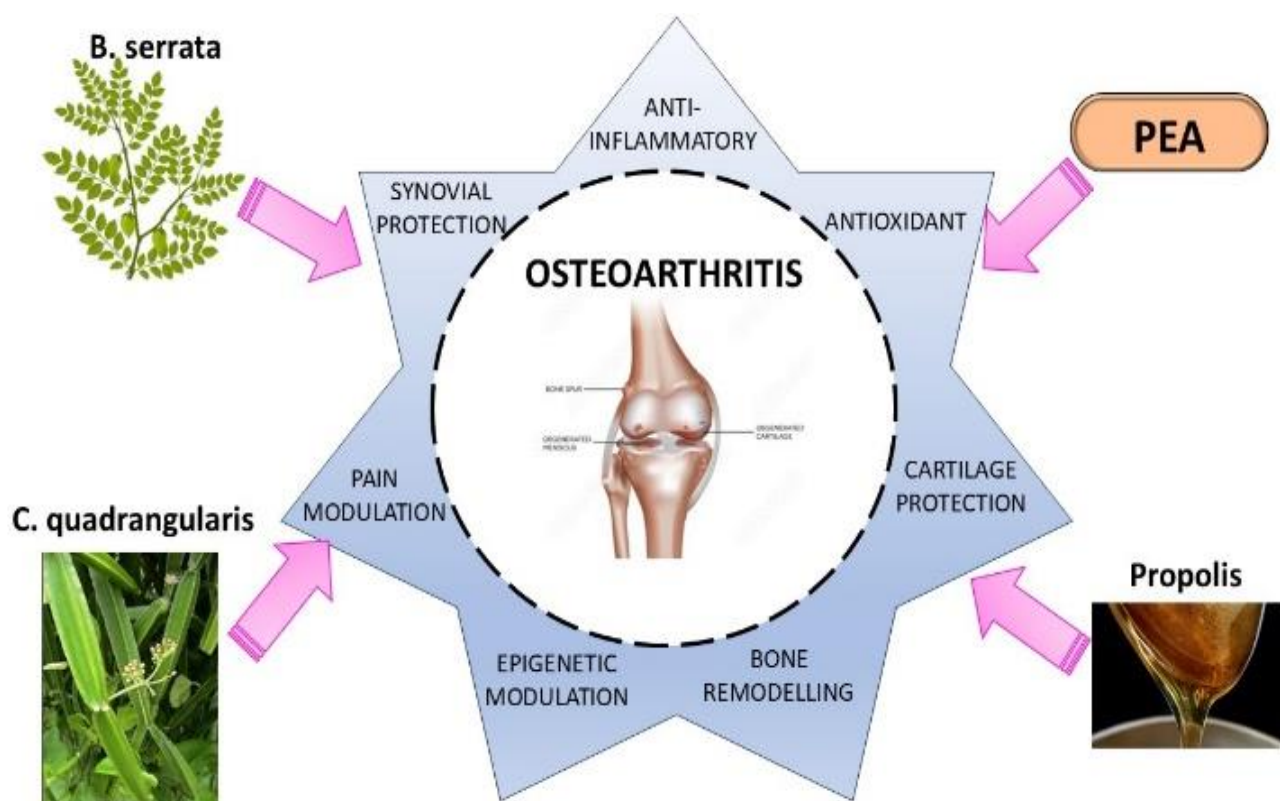


Fig. 1. Proposed drug combination.

**Table 1.** Evidence-based critical analysis of the mechanism of formulation

Mechanism of action	Combination of components	Individual components effects	References
Anti-inflammatory action	<i>B. serrata</i> , PEA, propolis	<i>B. serrata</i> : blocks 5-LOX and decreases LT production	[67–71]
		PEA: induces PPAR- $\alpha$ and decreases pro-inflammatory cytokines	[72–76]
		Propolis: alleviates NF- $\kappa$ B pathway and reduces IL-6, IL-11-1B, TNF- $\alpha$	[77–81]
ECM and Cartilage preservation	<i>B. serrata</i> , <i>C. quadrangularis</i>	<i>B. serrata</i> : inhibits MMP and protects cartilage from degradation	[82–86]
		<i>C. quadrangularis</i> : enhances synthesis of glycosaminoglycans and collagen, increases chondrocyte proliferation	[87–90]
Decrease in oxidative stress	Propolis, <i>B. serrata</i> , PEA	Propolis: improves SOD, catalase and glutathione peroxidase activities	[91–95]
		<i>B. serrata</i> : suppresses lipid peroxidation and scavenges free radicals	[96–99]
		PEA: improves antioxidant defense mechanisms	[100–103]
Modification of pain signalling pathways	PEA, <i>B. serrata</i> , propolis	PEA: decreases mast cells degranulation and prevents TRPV1 receptor activation	[104–108]
		<i>B. serrata</i> : prevents PKA, AKAP and PDE4 signalling to decrease nociception	[109–113]
		Propolis: modifies cannabinoid receptors and decreases substance P expression	[114–117]
Remodeling and regeneration of bone	<i>C. quadrangularis</i> , <i>B. serrata</i>	<i>C. quadrangularis</i> : improves differentiation and mineralization of osteoblast, enhances expression of RUNX2 and Osteopontin	[118–122]
		<i>B. serrata</i> : decreases the activity of osteoclast through inhibition of RANKL pathway	[123,124]
Protection of synovial membrane	PEA, propolis, <i>B. serrata</i>	PEA: suppresses inflammation and hyperplasia of synovium	[125–128]
		Propolis: suppresses activation and invasion of synovial fibroblast	[129–132]
		<i>B. serrata</i> : prevents synovial angiogenesis and inhibits synovial cellular infiltration	[133–136]
Modulation of gut-joint axis	Propolis, PEA	Propolis: improves the integrity of gut barrier and decreases microbial dysbiosis	[137–140]
		PEA: modifies the composition of gut microbiota and suppresses systemic inflammation	[141–144]
Epigenetic regulation	<i>B. serrata</i> , <i>C. quadrangularis</i>	<i>B. serrata</i> : controls histone deacetylases and patterns of DNA methylation in chondrocytes	[145–147]
		<i>C. quadrangularis</i> : modifies miRNA expression involved in maintaining cartilage homeostasis	[148–151]

#### 4.2. *B. serrata*

*B. serrata* is a medicinal plant, which is otherwise known as frankincense, and has been used for treating many inflammatory and musculoskeletal conditions. It has boswellic acids as bioactive compounds, and AKBA is the most efficient compound in preventing degradation of cartilage and inhibiting inflammatory pathways [54]. It has demonstrated inhibition of 5-lipoxygenase (5-LOX) and suppresses further production of leukotrienes [55]. Also, it modifies the NF- $\kappa$ B signalling, thereby decreasing the pro-inflammatory cytokines' expression. It was found to inhibit cathepsin G and prostaglandin E synthase-1 [56]. It was found to significantly reduce pain and improve physical function [57]. It was shown to have disease-modifying effects, which were found by decreased levels of MMP-3, a marker of cartilage destruction [58]. Studies have shown that it prevents apoptosis of chondrocytes and improves cartilage integrity [59].

#### 4.3. Propolis

It is a natural resin derived from bees that is rich in polyphenols, with anti-inflammatory and antioxidant effects [60]. It was found to decrease pro-inflammatory cytokines by inhibiting activation of NF- $\kappa$ B and reduce

joint inflammation by decreasing prostaglandins derived from COX-2 [61, 62]. It can reduce MMP-13 and production of nitric oxide (NO), thereby suppressing oxidative damage [63]. Thereby, it protects cartilage against wear and tear.

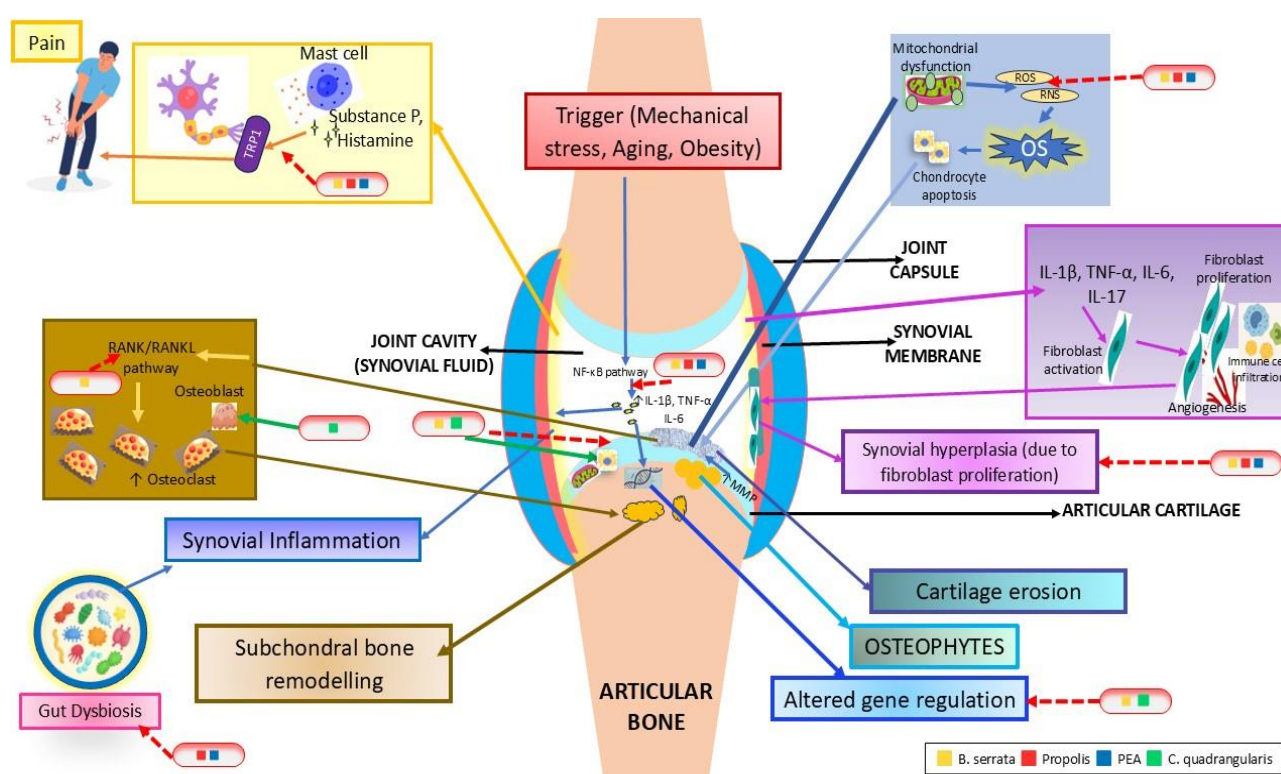
#### 4.4. PEA

PEA is known as an endogenous fatty acid amide that has important roles like anti-inflammatory, neuroprotective, and analgesic [64]. It acts by inducing PPAR- $\alpha$ , thereby reducing pro-inflammatory genes and consequently suppressing cytokine release. It also modulates mast cells, thereby preventing their degranulation and histamine-mediated inflammation in synovial tissues [65]. Also, it helps to decrease glial cell activation and controls neurogenic inflammation, thereby decreasing chronic pain. It was reported to interact with TRPV1 channels and cannabinoid receptors, further contributing to analgesia [66].

In this novel formulation, each ingredient targets multiple interconnected pathways in the pathophysiology of OA. The consolidated mechanisms provide anti-inflammatory, anti-oxidant, cartilage protection and pain modulation effects. Table 2 shows the comparison of the existing OA treatment with the current formulation. Hence, this formulation holds the potential to offer long-term benefit in OA.

**Table 2.** Comparison of the existing OA treatment with the new formulation.

Parameters	Existing OA treatment	New evidence-based formulation	References
Examples/ Composition	NSAIDs like diclofenac, ibuprofen, corticosteroids like prednisolone, Intra-articular hyaluronic acid injection, tramadol	<i>Cissus quadrangularis</i> , <i>Boswellia serrata</i> , propolis, palmitoylethanolamide (PEA)	[152–154]
Mechanism of action	Symptomatic treatment and suppressing inflammation	Multi-dimensional: Anti-inflammatory, antioxidant, matrix regeneration, cartilage protection and pain reduction	[152–154]
Inflammatory marker reduction	Moderate	Significant, as it targets multiple pathways	[155,156]
Cartilage protection	Mild to potentially destructive	Potential regenerative and protective effects	[157–159]
Safety profile	High risk of adverse effects	Minimal side effects	[160–162]
Long-term use	Not suitable, because of risk of organ toxicity	Promising tissue preservation that makes convenient long-term use and can be formulated in newer drug delivery designs	[163–165]



**Fig. 2.** Combined mechanistic action of the components of the drug. **Abbreviations:** OA – Osteoarthritis; PEA – palmitoylethanolamide; IL-1 $\beta$  – Interleukin 1 $\beta$ , NF- $\kappa$ B – Nuclear Factor kappa-light-chain-enhancer of activated B cells; IL-6 – Interleukin 6; IL-17 – Interleukin-17; MMPs – Matrix Metalloproteinases; RANK-Receptor Activator of Nuclear factor  $\kappa$ B; RANKL – RANK Ligand; RNS – Reactive Nitrogen Species; ROS – Reactive Oxygen Species; TNF- $\alpha$  – Tumor Necrosis Factor-alpha; TRPV1 – Transient Receptor Potential Vanilloid 1.

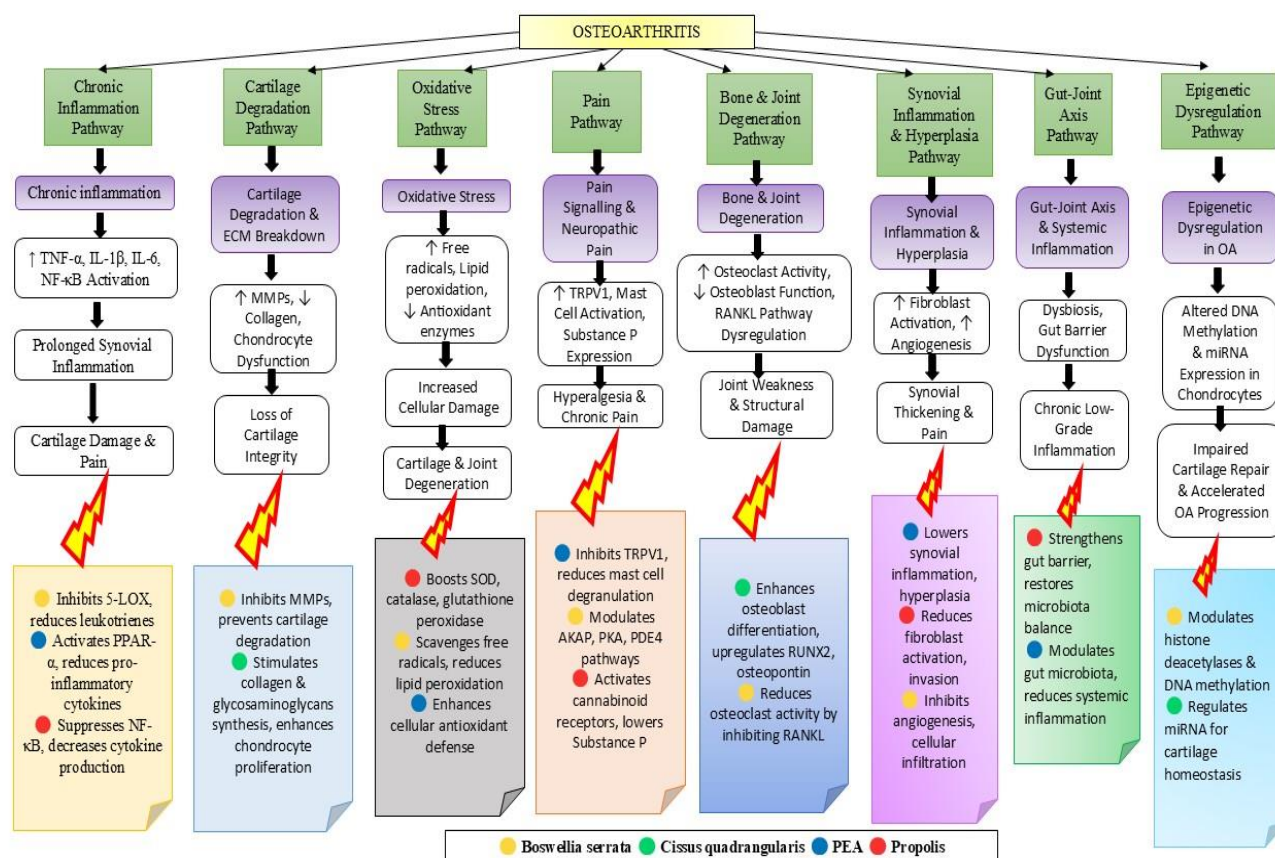
## 5. Discussion

As depicted in Fig. 2, the pathogenesis of OA is triggered and maintained by obesity, mechanical stress and aging, that drives oxidative stress and mitochondrial dysfunction.

Furthermore, Fig. 3 illustrates a wider integrated perspective on how this conceptual combination focuses diverse yet interconnected components of OA pathophysiology, comprising oxidative stress, chronic inflammation, cartilage degradation, synovial inflammation, pain signalling, epigenetic dysregulation, and gut-joint axis dysfunction. The pathways involved in the pathophysiology are chronic inflammation, oxidative stress, cartilage

degradation, bone and joint degradation, pain signalling, gut-joint axis dysfunction, synovial inflammation, and epigenetic dysregulation. *B. serrata* (Yellow code) suppresses inflammatory mediators. *C. quadrangularis* (Green code) prevents degradation of cartilage, improves chondrocyte proliferation, and supports collagen synthesis. PEA (Blue code) modulates pain signalling and reduces neurogenic inflammation. propolis (Red code) controls gut microbiota and alleviates systemic inflammation in OA. Together, these compounds are hypothesized to provide a multidimensional strategy to treat OA by its anti-inflammatory, antioxidant, cartilage-integrity- improving, matrix regeneration, and pain reduction properties based on existing scientific evidence.





**Fig. 3.** Combined mechanistic action of the components of the drug. This infographic depicts the interrelated pathways that contribute to the pathophysiology of OA and the specific actions of *B. serrata*, *C. quadrangularis*, PEA and propolis in targeting these pathways to execute therapeutic effects in OA.

## 6. Challenges and limitations

From the current review, it is proposed that this novel formulation might have a promising synergistic potential but with several limitations which need proper consideration. Firstly, this formulation is purely conceptual, based on existing preclinical and clinical data for the individual components of the combination without any direct scientific evidence for the combination as a whole. Secondly, the natural extracts of these individual components have potential variability in terms of bioavailability, quality, and standardization that could significantly affect formulation and thereby pose a regulatory challenge. Also, other aspects of this combination formulation on long-term safety and drug interactions are yet to be explored. These limitations emphasize the need for a large-scale clinical trial on this formulation for robust clinical validation and clinical adoption.

## 7. Future directions

In future, large-scale clinical trials must be conducted to properly validate its clinical efficacy, safety profile, and long-term benefits. Effective pharmaceutical formulation is crucial to ensure its efficient therapeutic outcomes. Table 3 represents the key aspects of pharmaceutical formulation. Additionally, its potential role in other phenotypes of OA must be investigated properly to validate its clinical utility. Proper regulatory standardization is mandatory for its

successful translation into a rationale-based, commercially possible OA treatment. Based on the promising mechanistic synergy and scientific support for the individual components, further studies are needed to explore its development, efficacy, stability, and safety. The authors are open to further translational research partnerships or academic collaboration to further explore and validate its potential as a novel treatment option for OA.

## 8. Purpose of this review

The current review was proposed in the context of an academic initiative to appraise thoroughly the possible synergistic potential of *C. quadrangularis*, *B. Serrata*, propolis and PEA in specifically targeting the multifaceted pathophysiology of OA. Even though the proposed formulation has not yet been evaluated clinically, it is entirely built upon existing robust preclinical and clinical studies on mechanistic evidence. The principal goal of this review article is not to recommend the proposed formulation for immediate clinical application, but rather to offer a scientifically sound rational basis for the purpose of further translational research and robust evidence-based drug development. Through presenting an evidence-based novel formulation strategy, the authors aim for future clinical translation of this conceptual formulation by further research collaboration and clinical validation.

**Table 3.** Key aspects of pharmaceutical formulation.

Key aspects	Analysis
Bioavailability challenges and solutions	<ul style="list-style-type: none"> <li><i>B. serrata</i>: Phospholipid complexation could improve the bioavailability of boswellic acid by 7-fold.</li> <li>Propolis: Liposomal encapsulation might improve flavonoid bioavailability.</li> <li>PEA: Micronization might increase the bioavailability and efficacy of PEA.</li> </ul>
Standardization	<ul style="list-style-type: none"> <li>Standardization is mandatory to ensure consistent level of active bioactive compounds like AKBA (&gt;30%) in <i>B. serrata</i>, flavonoid/ phenolic content in <i>propolis</i> and ketosteroid in <i>C. quadrangularis</i>.</li> </ul> <p>Modern analytical methods like spectroscopy, HPLC and LC-MS/MS can ensure its effective concentration.</p>
Stability	<ul style="list-style-type: none"> <li>To develop a tablet with good shelf-life, stability and consistent release profiles of active compounds, it is essential to do suitable excipient selection, compression properties and stability analysis.</li> <li>Hydroxypropyl Methylcellulose (HPMC) might improve controlled release of polyphenolic compounds.</li> </ul> <p>A thorough stability analysis must be done to assess any possible interactions among active compounds and to assess oxidative degradation of phenolic compounds through standard stability testing.</p>
Drug interactions	<ul style="list-style-type: none"> <li>Propolis components have been suggested to influence immunomodulatory pathways by few studies; that caution might be warranted in patients on immunosuppressive therapies.</li> <li><i>B. serrata</i> may enhance anticoagulants and hence should be used in caution with aspirin or warfarin.</li> <li>PEA has low drug interaction risk but has theoretical interaction with some CNS depressants.</li> <li>Intra-formulation interactions: Presence of antioxidant polyphenols in propolis could theoretically compete with PEA for metabolism and the added anti-inflammatory effects may improve efficacy but could also intensify immune modulation or mild GI side effects.</li> </ul>
Possible formulation type	<ul style="list-style-type: none"> <li>Possibly given as once-a-day oral tablet.</li> <li>Possible oral delivery formats are capsule, tablet or softgel.</li> <li>The choice depends upon target release kinetics, stability and bioavailability.</li> </ul>
Dosing strategy	<ul style="list-style-type: none"> <li><i>B. serrata</i>: 100–250 mg/ day of standardized <i>Boswellia</i> extract.</li> <li><i>C. quadrangularis</i>: 500–1000 mg/day of standardized ketosteroid containing cissus extract.</li> <li>PEA: 300–600 mg/day of micronized PEA.</li> <li>Propolis: 250–500 mg/day of standardized propolis extract.</li> </ul>
Target patient group	<ul style="list-style-type: none"> <li>The potential target patient group include inflammatory OA phenotype, adjuvant OA therapy, patients contraindicated to NSAIDs.</li> <li>It has a strong conceptual potential to be used as a treatment of OA, as a part of early intervention.</li> </ul>

## 9. Conclusions

This novel conceptual formulation, consisting of *B. serrata*, *C. quadrangularis*, PEA, and propolis provides a multi-targeted treatment strategy for OA, as it addresses various pathophysiological pathways in OA such as oxidative stress, chronic inflammation, cartilage degradation, and pain signalling. In contrast to conventional treatment options, which primarily provide symptom relief, this novel formulation could offer analgesic, chondroprotective, and regenerative effects with a reasonable safety profile, making it suitable for long-term use. The overall effects of this formulation could make it a potential drug to treat OA, especially in patients with inflammatory OA and those contraindicated for conventional NSAIDs. Based on existing evidence about treating OA by individual components from various studies on their rationale and safety profile, this current review suggests that this formulation has a strong translational potential.

**Author Contributions:** Conceptualization, Dr.B.Dharani and Dr.Suba.A.; methodology, Dr.B.Dharani.; validation, Dr.B.Dharani and Dr.Suba.A.; investigation, Dr.B.Dharani; resources, Dr.Suba.A.; data curation, Dr.B.Dharani; writing—original draft preparation, Dr.B.Dharani; writing—review and editing, Dr.Suba.A.; visualization, Dr.B.Dharani; supervision, Dr.Suba.A.; project administration, Dr.B.Dharani. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research received no external funding.

**Acknowledgments:** We express our sincere thanks to

all the scientists and researchers whose invaluable contribution has serves as the major pillar for this research. The authors conceptualized this novel formulation as a part of an academic initiative to develop evidence-based combination formulation for management of OA. To our knowledge, this is the first review article to explore this combination. This formulation is presented for academic discussion and exploration for future translation and the authors welcome further research collaboration and development inquiries.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

- GBD 2017 Disease and Injury Incidence and Prevalence Collaborators Global, Regional, and National Incidence, Prevalence, and Years Lived with Disability for 354 Diseases and Injuries for 195 Countries and Territories, 1990-2017: A Systematic Analysis for the Global Burden of Disease Study 2017. *Lancet* **2018**, 392, 1789-1858. DOI: 10.1016/S0140-6736(18)32279-7
- Clynes, M.A.; Jameson, K.A.; Edwards, M.H.; Cooper, C.; Dennison, E.M. Impact of Osteoarthritis on Activities of Daily Living: Does Joint Site Matter? *Aging Clin. Exp. Res.* **2019**, 31, 1049-1056. DOI: 10.1007/s40520-019-01163-0
- Ranjithkumar, N.; Paul, J.; Alagesan, J.; Viswanathan, R. Comparative Effectiveness of Extracorporeal Short Wave Therapy, Low-Level Laser Therapy, and Ultrasound in the Treatment of Rotator Cuff Tendinopathy. *Biomed. Pharmacol. J.* **2025**, 18, 849-866. doi: 10.13005/bpj/3134

4. Oleshchuk, O.; Pinyazhko, O.; Klantsa, M.; Posokhova, K.; Lukanyuk, M.; Mahanova, T.; Shanida, M. Critical Assessment of Effectiveness and Safety of Tramadol and Evaluation of Its Market in Ukraine. *Biomed. Pharmacol. J.* **2024**, *17*, 2087-2109. DOI: 10.13005/bpj/3010
5. Fu, K.; Si, S.; Jin, X.; Zhang, Y.; Duong, V.; Cai, Q.; Li, G.; Oo, W.M.; Zheng, X.; Boer, C.G.; et al. Exploring Antidiabetic Drug Targets as Potential Disease-Modifying Agents in Osteoarthritis. *EBioMedicine* **2024**, *107*, Art. No: 105285. DOI: 10.1016/j.ebiom.2024.105285
6. Li, S.; Cao, P.; Chen, T.; Ding, C. Latest Insights in Disease-Modifying Osteoarthritis Drugs Development. *Ther. Adv. Musculoskelet. Dis.* **2023**, *15*, Art. No: 1759720X231169839. DOI: 10.1177/1759720X231169839
7. Kim, H.; Seo, J.; Lee, Y.; Park, K.; Perry, T.A.; Arden, N.K.; Mobasheri, A.; Choi, H. The Current State of the Osteoarthritis Drug Development Pipeline: A Comprehensive Narrative Review of the Present Challenges and Future Opportunities. *Ther. Adv. Musculoskelet. Dis.* **2022**, *14*, Art. No: 1759720X221085952. DOI: 10.1177/1759720X221085952
8. Maouche, A.; Boumediene, K.; Baugé, C. Bioactive Compounds in Osteoarthritis: Molecular Mechanisms and Therapeutic Roles. *Int. J. Mol. Sci.* **2024**, *25*, Art. No: 11656. DOI: 10.3390/ijms252111656
9. Azam, Z.; Sapra, L.; Baghel, K.; Sinha, N.; Gupta, R.K.; Soni, V.; Saini, C.; Mishra, P.K.; Srivastava, R.K. Cissus Quadrangularis (Hadjod) Inhibits RANKL-Induced Osteoclastogenesis and Augments Bone Health in an Estrogen-Deficient Preclinical Model of Osteoporosis via Modulating the Host Osteoimmune System. *Cells* **2023**, *12*, Art. No: 216. DOI: 10.3390/cells12020216
10. Nath, R.; Kar, B.K.; Dhadiwal, R.K.; Daftary, G.V.; Khemnari, B.M.; Patil, N.N. Role of Cissus Quadrangularis in Bone Loss Pathologies. *Int. J. Orthop. Sci.* **2024**, *10*, 196-201.
11. Alluri, V.K.; Kundimi, S.; Sengupta, K.; Golakoti, T.; Kilari, E.K. An Anti-Inflammatory Composition of Boswellia Serrata Resin Extracts Alleviates Pain and Protects Cartilage in Monoiodoacetate-Induced Osteoarthritis in Rats. *Evid. Based. Complement. Alternat. Med.* **2020**, *2020*, Art. No: 7381625. DOI: 10.1155/2020/7381625
12. Kurek-Górecka, A.; Rzepecka-Stojko, A.; Górecki, M.; Stojko, J.; Sosada, M.; Swierczek-Zieba, G. Structure and Antioxidant Activity of Polyphenols Derived from Propolis. *Molecules* **2013**, *19*, 78-101. DOI: 10.3390/molecules19010078
13. Branković, M.; Gmizić, T.; Dukić, M.; Zdravković, M.; Daskalović, B.; Mrda, D.; Nikolić, N.; Brajković, M.; Gojčić, M.; Lalatović, J.; et al. Therapeutic Potential of Palmitoylethanolamide in Gastrointestinal Disorders. *Antioxidants (Basel)* **2024**, *13*, Art. No: 600. DOI: 10.3390/antiox13050600
14. Costa, B.; Comelli, F.; Bettoni, I.; Colleoni, M.; Giagnoni, G. The Endogenous Fatty Acid Amide, Palmitoylethanolamide, Has Anti-Allodynic and Anti-Hyperalgesic Effects in a Murine Model of Neuropathic Pain: Involvement of CB(1), TRPV1 and PPARgamma Receptors and Neurotrophic Factors. *Pain* **2008**, *139*, 541-550. DOI: 10.1016/j.pain.2008.06.003
15. Yao, Q.; Wu, X.; Tao, C.; Gong, W.; Chen, M.; Qu, M.; Zhong, Y.; He, T.; Chen, S.; Xiao, G. Osteoarthritis: Pathogenic Signaling Pathways and Therapeutic Targets. *Signal Transduct. Target. Ther.* **2023**, *8*, Art. No: 56. DOI: 10.1038/s41392-023-01330-w
16. Yunus, M.H.M.; Nordin, A.; Kamal, H. Pathophysiological Perspective of Osteoarthritis. *Medicina (Kaunas)* **2020**, *56*, Art. No: 614. DOI: 10.3390/medicina56110614
17. Berenbaum, F. Osteoarthritis as an Inflammatory Disease (Osteoarthritis Is Not Osteoarthritis!). *Osteoarthritis Cartilage* **2013**, *21*, 16-21. DOI: 10.1016/j.joca.2012.11.012
18. Wojdasiewicz, P.; Poniatowski, Ł.A.; Szukiewicz, D. The Role of Inflammatory and Anti-Inflammatory Cytokines in the Pathogenesis of Osteoarthritis. *Mediators Inflamm.* **2014**, *2014*, Art. No: 561459. DOI: 10.1155/2014/561459
19. Zheng, L.; Zhang, Z.; Sheng, P.; Mobasheri, A. The Role of Metabolism in Chondrocyte Dysfunction and the Progression of Osteoarthritis. *Ageing Res. Rev.* **2021**, *66*, Art. No: 101249. DOI: 10.1016/j.arr.2020.101249
20. Chen, C.; Xie, J.; Rajappa, R.; Deng, L.; Fredberg, J.; Yang, L. Interleukin-18 and Tumor Necrosis Factor- $\alpha$  Increase Stiffness and Impair Contractile Function of Articular Chondrocytes. *Acta Biochim. Biophys. Sin. (Shanghai)* **2015**, *47*, 121-129. DOI: 10.1093/abbs/gmu116
21. Pratta, M.A.; Yao, W.; Decicco, C.; Tortorella, M.D.; Liu, R.-Q.; Copeland, R.A.; Magolda, R.; Newton, R.C.; Trzaskos, J.M.; Arner, E.C. Aggrecan Protects Cartilage Collagen from Proteolytic Cleavage. *J. Biol. Chem.* **2003**, *278*, 45539-45545. DOI: 10.1074/jbc.M303737200
22. Akkiraju, H.; Nohe, A. Role of Chondrocytes in Cartilage Formation, Progression of Osteoarthritis and Cartilage Regeneration. *J. Dev. Biol.* **2015**, *3*, 177-192. DOI: 10.3390/jdb3040177
23. Pulik, Ł.; Łęgosz, P.; Motyl, G. Matrix Metalloproteinases in Rheumatoid Arthritis and Osteoarthritis: A State of the Art Review. *Reumatologia* **2023**, *61*, 191-201. DOI: 10.5114/reum/168503
24. Tajdari, M.; Peyrovinasab, A.; Bayanati, M.; Ismail Mahboubi Rabbani, M.; Abdolghaffari, A.H.; Zarghi, A. Dual COX-2/TNF- $\alpha$  Inhibitors as Promising Anti-Inflammatory and Cancer Chemopreventive Agents: A Review. *Iran. J. Pharm. Res.* **2024**, *23*, Art. No: e151312. DOI: 10.5812/ijpr-151312
25. Wang, Y.; Che, M.; Xin, J.; Zheng, Z.; Li, J.; Zhang, S. The Role of IL-18 and TNF- $\alpha$  in Intervertebral Disc Degeneration. *Biomed. Pharmacother.* **2020**, *131*, Art. No: 110660. DOI: 10.1016/j.biopha.2020.110660
26. Robinson, W.H.; Lepus, C.M.; Wang, Q.; Raghu, H.; Mao, R.; Lindstrom, T.M.; Sokolove, J. Low-Grade Inflammation as a Key Mediator of the Pathogenesis of Osteoarthritis. *Nat. Rev. Rheumatol.* **2016**, *12*, 580-592. DOI: 10.1038/nrrheum.2016.136
27. Link, T.M.; Li, X. Establishing Compositional MRI of Cartilage as a Biomarker for Clinical Practice. *Osteoarthritis Cartilage* **2018**, *26*, 1137-1139. DOI: 10.1016/j.joca.2018.02.902



28. Lepetsos, P.; Papavassiliou, A.G. ROS/Oxidative Stress Signaling in Osteoarthritis. *Biochim. Biophys. Acta* **2016**, *1862*, 576-591. DOI: 10.1016/j.bbadis.2016.01.003
29. Ansari, M.Y.; Ahmad, N.; Haqqi, T.M. Oxidative Stress and Inflammation in Osteoarthritis Pathogenesis: Role of Polyphenols. *Biomed. Pharmacother.* **2020**, *129*, Art. No: 110452. DOI: 10.1016/j.biopha.2020.110452
30. Bolduc, J.A.; Collins, J.A.; Loeser, R.F. Reactive Oxygen Species, Aging and Articular Cartilage Homeostasis. *Free Radic. Biol. Med.* **2019**, *132*, 73-82. DOI: 10.1016/j.freeradbiomed.2018.08.038
31. Guo, P.; Alhaskawi, A.; Adel Abdo Moqbel, S.; Pan, Z. Recent Development of Mitochondrial Metabolism and Dysfunction in Osteoarthritis. *Front. Pharmacol.* **2025**, *16*, Art. No: 1538662. DOI: 10.3389/fphar.2025.1538662
32. Li, S.; Xiong, Y.; Zhu, H.; Ma, T.; Sun, X.; Xiao, J. Microenvironment-Responsive Nanosystems for Osteoarthritis Therapy. *Engineered Regeneration* **2024**, *5*, 92-110. DOI: 10.1016/j.engreg.2023.12.002
33. Kim, J.-H.; Jeon, J.; Shin, M.; Won, Y.; Lee, M.; Kwak, J.-S.; Lee, G.; Rhee, J.; Ryu, J.-H.; Chun, C.-H.; et al. Regulation of the Catabolic Cascade in Osteoarthritis by the Zinc-ZIP8-MTF1 Axis. *Cell* **2014**, *156*, 730-743. DOI: 10.1016/j.cell.2014.01.007
34. Fox, S.; Bedi, A.J.; Rodeo, A. The Basic Science of Articular Cartilage: Structure, Composition, and Function. *Sports Health* **2009**, *1*, 461-468.
35. Troeberg, L.; Nagase, H. Proteases Involved in Cartilage Matrix Degradation in Osteoarthritis. *Biochim. Biophys. Acta* **2012**, *1824*, 133-145. DOI: 10.1016/j.bbapap.2011.06.020
36. Ur Rehman, S.; Iqbal, S.; Shahid, U.; Jahangir, S.; Malik, L. Cartilage: Structure, Function, and the Pathogenesis of Osteoarthritis. In *Advancements in Synovial Joint Science - Structure, Function, and Beyond. IntechOpen*; **2024**.
37. Henao-Murillo, L.; Pastrama, M.-I.; Ito, K.; van Donkelaar, C.C. The Relationship between Proteoglycan Loss, Overloading-Induced Collagen Damage, and Cyclic Loading in Articular Cartilage. *Cartilage* **2021**, *13*, 1501S-1512S. DOI: 10.1177/1947603519885005
38. Mukherjee, A.; Das, B. The Role of Inflammatory Mediators and Matrix Metalloproteinases (MMPs) in the Progression of Osteoarthritis. *Biomater. Biosyst.* **2024**, *13*, Art. No: 100090. DOI: 10.1016/j.bbiosy.2024.100090
39. Sanchez-Lopez, E.; Coras, R.; Torres, A.; Lane, N.E.; Guma, M. Synovial Inflammation in Osteoarthritis Progression. *Nat. Rev. Rheumatol.* **2022**, *18*, 258-275. DOI: 10.1038/s41584-022-00749-9
40. Mathiessen, A.; Conaghan, P.G. Synovitis in Osteoarthritis: Current Understanding with Therapeutic Implications. *Arthritis Res. Ther.* **2017**, *19*, Art. No: 18. DOI: 10.1186/s13075-017-1229-9
41. Schaible, H.-G.; Ebersberger, A.; Natura, G. Update on Peripheral Mechanisms of Pain: Beyond Prostaglandins and Cytokines. *Arthritis Res. Ther.* **2011**, *13*, Art. No: 210. DOI: 10.1186/ar3305.
42. Eitner, A.; Hofmann, G.O.; Schaible, H.-G. Mechanisms of Osteoarthritic Pain. Studies in Humans and Experimental Models. *Front. Mol. Neurosci.* **2017**, *10*, Art. No: 349. DOI: 10.3389/fnmol.2017.00349
43. Volcheck, M.M.; Graham, S.M.; Fleming, K.C.; Mohabbat, A.B.; Luedtke, C.A. Central Sensitization, Chronic Pain, and Other Symptoms: Better Understanding, Better Management. *Cleve. Clin. J. Med.* **2023**, *90*, 245-254. DOI: 10.3949/ccjm.90a.22019
44. Wen, B.; Pan, Y.; Cheng, J.; Xu, L.; Xu, J. The Role of Neuroinflammation in Complex Regional Pain Syndrome: A Comprehensive Review. *J. Pain Res.* **2023**, *16*, 3061-3073. DOI: 10.2147/JPR.S423733
45. Yang, D.; Xu, J.; Xu, K.; Xu, P. Skeletal Interoception in Osteoarthritis. *Bone Res.* **2024**, *12*, Art. No: 22. DOI: 10.1038/s41413-024-00328-6
46. Hu, Y.; Chen, X.; Wang, S.; Jing, Y.; Su, J. Subchondral Bone Microenvironment in Osteoarthritis and Pain. *Bone Res.* **2021**, *9*, 20, DOI: 10.1038/s41413-021-00147-z
47. Zhu, S.; Zhu, J.; Zhen, G.; Hu, Y.; An, S.; Li, Y.; Zheng, Q.; Chen, Z.; Yang, Y.; Wan, M.; et al. Subchondral Bone Osteoclasts Induce Sensory Innervation and Osteoarthritis Pain. *J. Clin. Invest.* **2019**, *129*, 1076-1093. DOI: 10.1172/JCI121561
48. Singh, P.; Gupta, A.; Qayoom, I.; Singh, S.; Kumar, A. Orthobiologics with Phytobioactive Cues: A Paradigm in Bone Regeneration. *Biomed. Pharmacother.* **2020**, *130*, Art. No: 110754. DOI: 10.1016/j.biopha.2020.110754
49. Mishra, G.; Srivastava, S.; Nagori, B.P. Pharmacological and Therapeutic Activity of *Cissus Quadrangularis*: An Overview. *Int. J. Pharm. Tech. Res.* **2010**, *2*, 1298-1310.
50. Bhujade, A.M.; Talmale, S.; Kumar, N.; Gupta, G.; Reddanna, P.; Das, S.K.; Patil, M.B. Evaluation of *Cissus Quadrangularis* Extracts as an Inhibitor of COX, 5-LOX, and Proinflammatory Mediators. *J. Ethnopharmacol.* **2012**, *141*, 989-996. DOI: 10.1016/j.jep.2012.03.044
51. Bloomer, R.J.; Farney, T.M.; McCarthy, C.G.; Lee, S.-R. *Cissus Quadrangularis* Reduces Joint Pain in Exercise-Trained Men: A Pilot Study. *Phys. Sportsmed.* **2013**, *41*, 29-35. DOI: 10.3810/psm.2013.09.2021
52. Kanwar, J.; Samarasinghe, R.; Kumar, K.; Arya, R.; Sharma, S.; Zhou, S.-F.; Sasidharan, S.; Kanwar, R. *Cissus Quadrangularis* Inhibits IL-1 $\beta$  Induced Inflammatory Responses on Chondrocytes and Alleviates Bone Deterioration in Osteotomized Rats via P38 MAPK Signaling [Corrigendum]. *Drug Des. Devel. Ther.* **2017**, *11*, 2683-2684. DOI: 10.2147/dddt.s148615
53. Banu, J.; Varela, E.; Bahadur, A.N.; Soomro, R.; Kazi, N.; Fernandes, G. Inhibition of Bone Loss by *Cissus Quadrangularis* in Mice: A Preliminary Report. *J. Osteoporos.* **2012**, *2012*, Art. No: 101206. DOI: 10.1155/2012/101206
54. Iram, F.; Khan, S.A.; Husain, A. Phytochemistry and Potential Therapeutic Actions of Boswellic Acids: A Mini-Review. *Asian Pac. J. Trop. Biomed.* **2017**, *7*, 513-523. DOI: 10.1016/j.apjtb.2017.05.001
55. Siddiqui, M.Z. Boswellia Serrata, a Potential Antiinflammatory Agent: An Overview. *Indian J. Pharm. Sci.* **2011**, *73*, 255-261. DOI: 10.4103/0250-474X.93507

56. Sengupta, K.; Alluri, K.V.; Satish, A.R.; Mishra, S.; Golakoti, T.; Sarma, K.V.; Dey, D.; Raychaudhuri, S.P. A Double Blind, Randomized, Placebo Controlled Study of the Efficacy and Safety of 5-Loxin for Treatment of Osteoarthritis of the Knee. *Arthritis Res. Ther.* **2008**, *10*, Art. No: R85. DOI: 10.1186/ar2461
57. Yu, G.; Xiang, W.; Zhang, T.; Zeng, L.; Yang, K.; Li, J. Effectiveness of Boswellia and Boswellia Extract for Osteoarthritis Patients: A Systematic Review and Meta-Analysis. *BMC Complement. Med. Ther.* **2020**, *20*, Art. No: 225. DOI: 10.1186/s12906-020-02985-6
58. Vishal, A.A.; Mishra, A.; Raychaudhuri, S.P. A Double Blind, Randomized, Placebo Controlled Clinical Study Evaluates the Early Efficacy of Aflapin in Subjects with Osteoarthritis of Knee. *Int. J. Med. Sci.* **2011**, *8*, 615-622. DOI: 10.7150/ijms.8.615
59. Sukhikh, S.; Noskova, S.; Ivanova, S.; Ulrikh, E.; Izgaryshev, A.; Babich, O. Chondroprotection and Molecular Mechanism of Action of Phytonutraceuticals on Osteoarthritis. *Molecules* **2021**, *26*, Art. No: 2391. DOI: 10.3390/molecules26082391
60. Hossain, R.; Quispe, C.; Khan, R.A.; Saikat, A.S.M.; Ray, P.; Ongalbek, D.; Yeskaliyeva, B.; Jain, D.; Smeriglio, A.; Trombetta, D.; et al. propolis: An Update on Its Chemistry and Pharmacological Applications. *Chin. Med.* **2022**, *17*, Art. No: 100. DOI: 10.1186/s13020-022-00651-2
61. Oršolić, N.; Jazvinščak Jembrek, M. Potential Strategies for Overcoming Drug Resistance Pathways Using propolis and Its Polyphenolic/Flavonoid Compounds in Combination with Chemotherapy and Radiotherapy. *Nutrients* **2024**, *16*, Art. No: 3741. DOI: 10.3390/nu16213741
62. Altabbal, S.; Athamnah, K.; Rahma, A.; Wali, A.F.; Eid, A.H.; Iratni, R.; Al Dhaheri, Y. propolis: A Detailed Insight of Its Anticancer Molecular Mechanisms. *Pharmaceuticals (Basel)* **2023**, *16*, Art. No: 450. DOI: 10.3390/ph16030450
63. Arias, C.; Vásquez, B.; Salazar, L.A. propolis as a Potential Therapeutic Agent to Counteract Age-Related Changes in Cartilage: An in Vivo Study. *Int. J. Mol. Sci.* **2023**, *24*, Art. No: 14272. DOI: 10.3390/ijms241814272
64. Petrosino, S.; Marzo, D. The Pharmacology of Palmitoylethanolamide and First Data on the Therapeutic Efficacy of Some of Its New Formulations: Palmitoylethanolamide and Its New Formulations. *Br. J. Pharmacol.* **2017**, *174*, 1349-1365.
65. Skaper, S.D.; Facci, L.; Fusco, M.; Della Valle, M.F.; Zusso, M.; Costa, B.; Giusti, P. Palmitoylethanolamide, a Naturally Occurring Disease-Modifying Agent in Neuropathic Pain. *Inflammopharmacology* **2014**, *22*, 79-94. DOI: 10.1007/s10787-013-0191-7
66. Gabrielsson, L.; Mattsson, S.; Fowler, C.J. Palmitoylethanolamide for the Treatment of Pain: Pharmacokinetics, Safety and Efficacy. *Br. J. Clin. Pharmacol.* **2016**, *82*, 932-942. DOI: 10.1111/bcp.13020
67. Sengupta, K.; Kolla, J.N.; Krishnaraju, A.V.; Yalamanchili, N.; Rao, C.V.; Golakoti, T.; Raychaudhuri, S.; Raychaudhuri, S.P. Cellular and Molecular Mechanisms of Anti-Inflammatory Effect of Aflapin: A Novel Boswellia Serrata Extract. *Mol. Cell. Biochem.* **2011**, *354*, 189-197. DOI: 10.1007/s11010-011-0818-1
68. Sengupta, K.; Krishnaraju, A.V.; Vishal, A.A.; Mishra, A.; Trimurtulu, G.; Sarma, K.V.S.; Raychaudhuri, S.K.; Raychaudhuri, S.P. Comparative Efficacy and Tolerability of 5-Loxin and Aflapin Against Osteoarthritis of the Knee: A Double Blind, Randomized, Placebo Controlled Clinical Study. *Int. J. Med. Sci.* **2010**, *7*, 366-377. DOI: 10.7150/ijms.7.366
69. Bannuru, R.R.; Osani, M.C.; Al-Eid, F.; Wang, C. Efficacy of Curcumin and Boswellia for Knee Osteoarthritis: Systematic Review and Meta-Analysis. *Semin. Arthritis Rheum.* **2018**, *48*, 416-429. DOI: 10.1016/j.semarthrit.2018.03.001
70. Umar, S.; Umar, K.; Sarwar, A.H.M.G.; Khan, A.; Ahmad, N.; Ahmad, S.; Katiyar, C.K.; Husain, S.A.; Khan, H.A. Boswellia Serrata Extract Attenuates Inflammatory Mediators and Oxidative Stress in Collagen Induced Arthritis. *Phytomedicine* **2014**, *21*, 847-856. DOI: 10.1016/j.phymed.2014.02.001
71. Riva, A.; Ronchi, M.; Petrangolini, G.; Bosio, S.; Allegrini, P. Improved Oral Absorption of Quercetin from Quercetin Phytosome®, a New Delivery System Based on Food Grade Lecithin. *Eur. J. Drug Metab. Pharmacokinet.* **2019**, *44*, 169-177. DOI: 10.1007/s13318-018-0517-3
72. Lang-Illievich, K.; Klivinyi, C.; Lasser, C.; Brenna, C.T.A.; Szilagyi, I.S.; Bornemann-Ciment, H. Palmitoylethanolamide in the Treatment of Chronic Pain: A Systematic Review and Meta-Analysis of Double-Blind Randomized Controlled Trials. *Nutrients* **2023**, *15*, Art. No: 1350. DOI: 10.3390/nu15061350
73. Steels, E.; Venkatesh, R.; Steels, E.; Vitetta, G.; Vitetta, L. A Double-Blind Randomized Placebo Controlled Study Assessing Safety, Tolerability and Efficacy of Palmitoylethanolamide for Symptoms of Knee Osteoarthritis. *Inflammopharmacology* **2019**, *27*, 475-485. DOI: 10.1007/s10787-019-00582-9
74. Jung, J.I.; Lee, H.S.; Jeon, Y.E.; Kim, S.M.; Hong, S.H.; Moon, J.M.; Lim, C.Y.; Kim, Y.H.; Kim, E.J. Anti-Inflammatory Activity of Palmitoylethanolamide Ameliorates Osteoarthritis Induced by Monosodium Iodoacetate in Sprague-Dawley Rats. *Inflammopharmacology* **2021**, *29*, 1475-1486. DOI: 10.1007/s10787-021-00870-3
75. Gugliandolo, E.; Fusco, R.; Biundo, F.; D'Amico, R.; Benedetto, F.; Di Paola, R.; Cuzzocrea, S. Palmitoylethanolamide and Polydatin Combination Reduces Inflammation and Oxidative Stress in Vascular Injury. *Pharmacol. Res.* **2017**, *123*, 83-92. DOI: 10.1016/j.phrs.2017.06.014
76. Marini, I.; Bartolucci, M.L.; Bortolotti, F.; Gatto, M.R.; Bonetti, G.A. Palmitoylethanolamide versus a Nonsteroidal Anti-Inflammatory Drug in the Treatment of Temporomandibular Joint Inflammatory Pain. *J. Orofac. Pain* **2012**, *26*, 99-104.
77. Berretta, A.A.; Silveira, M.A.D.; Córdor Capcha, J.M.; De Jong, D. propolis and Its Potential against SARS-CoV-2 Infection Mechanisms and COVID-19 Disease: Running Title: Propolis against SARS-CoV-2 Infection and COVID-19. *Biomed. Pharmacother.* **2020**, *131*, Art. No: 110622. DOI: 10.1016/j.biopha.2020.110622

78. Kurek-Górecka, A.; Górecki, M.; Rzepecka-Stojko, A.; Balwierz, R.; Stojko, J. Bee Products in Dermatology and Skin Care. *Molecules* **2020**, *25*, Art. No: 556. DOI: 10.3390/molecules25030556
79. Pahlavani, N.; Malekhamadi, M.; Firouzi, S.; Rostami, D.; Sedaghat, A.; Moghaddam, A.B.; Ferns, G.A.; Navashenaq, J.G.; Reazvani, R.; Safarian, M.; et al. Molecular and Cellular Mechanisms of the Effects of propolis in Inflammation, Oxidative Stress and Glycemic Control in Chronic Diseases. *Nutr. Metab. (Lond.)* **2020**, *17*, Art. No: 65. DOI: 10.1186/s12986-020-00485-5
80. Xuan, H.; Yuan, W.; Chang, H.; Liu, M.; Hu, F. Anti-Inflammatory Effects of Chinese propolis in Lipopolysaccharide-Stimulated Human Umbilical Vein Endothelial Cells by Suppressing Autophagy and MAPK/NF- $\kappa$ B Signaling Pathway. *Inflammopharmacology* **2019**, *27*, 561-571. DOI: 10.1007/s10787-018-0533-6
81. Zulhendri, F.; Lesmana, R.; Tandean, S.; Christoper, A.; Chandrasekaran, K.; Irsyam, I.; Suwantika, A.A.; Abdulah, R.; Wathoni, N. Recent Update on the Anti-Inflammatory Activities of propolis. *Molecules* **2022**, *27*, Art. No: 8473. DOI: 10.3390/molecules27238473
82. Majeed, M.; Majeed, S.; Narayanan, N.K.; Nagabhushanam, K. A Pilot, Randomized, Double-Blind, Placebo-Controlled Trial to Assess the Safety and Efficacy of a Novel Boswellia Serrata Extract in the Management of Osteoarthritis of the Knee: A Novel B. Serrata Extract for Knee Osteoarthritis. *Phytother. Res.* **2019**, *33*, 1457-1468.
83. Minorette, P.; Santiago Sáez, A.; Liaño Riera, M.; Gómez Serrano, M.; García Martín, Á. Efficacy and Safety of Two Chondroprotective Supplements in Patients with Knee Osteoarthritis: A Randomized, Single-Blind, Pilot Study. *Cureus* **2024**, *16*, Art. No: e57579. DOI: 10.7759/cureus.57579
84. Hussain, H.; Wang, D.; El-Seedi, H.R.; Rashan, L.; Ahmed, I.; Abbas, M.; Mamadalieva, N.Z.; Sultani, H.N.; Hussain, M.I.; Shah, S.T.A. Therapeutic Potential of Boswellic Acids: An Update Patent Review (2016-2023). *Expert Opin. Ther. Pat.* **2024**, *34*, 723-732. DOI: 10.1080/13543776.2024.2369626
85. Villalvilla, A.; da Silva, J.A.; Largo, R.; Gualillo, O.; Vieira, P.C.; Herrero-Baumont, G.; Gómez, R. 6-Shogaol Inhibits Chondrocytes' Innate Immune Responses and Cathepsin-K Activity. *Mol. Nutr. Food Res.* **2014**, *58*, 256-266. DOI: 10.1002/mnfr.201200833
86. Shin, M.-R.; Kim, H.-Y.; Choi, H.-Y.; Park, K.S.; Choi, H.J.; Roh, S.-S. Boswellia Serrata Extract, 5-Loxin®, Prevents Joint Pain and Cartilage Degeneration in a Rat Model of Osteoarthritis through Inhibition of Inflammatory Responses and Restoration of Matrix Homeostasis. *Evid. Based. Complement. Alternat. Med.* **2022**, *2022*, Art. No: 3067526. DOI: 10.1155/2022/3067526
87. Potu, B.K.; Bhat, K.M.R.; Rao, M.S.; Nampurath, G.K.; Chamallamudi, M.R.; Nayak, S.R.; Muttigi, M.S. Petroleum Ether Extract of *Cissus Quadrangularis* (Linn.) Enhances Bone Marrow Mesenchymal Stem Cell Proliferation and Facilitates Osteoblastogenesis. *Clinics (Sao Paulo)* **2009**, *64*, 993-998. DOI: 10.1590/S1807-59322009001000010
88. Bafna, P.S.; Patil, P.H.; Maru, S.K.; Mutha, R.E. *Cissus Quadrangularis* L: A Comprehensive Multidisciplinary Review. *J. Ethnopharmacol.* **2021**, *279*, Art. No: 114355. DOI: 10.1016/j.jep.2021.114355.
89. Sawangjit, R.; Puttarak, P.; Saokaew, S.; Chaikunapruk, N. Efficacy and Safety of *Cissus Quadrangularis* L. in Clinical Use: A Systematic Review and Meta-Analysis of Randomized Controlled Trials: Efficacy and Safety of *Cissus* in Clinical Use. *Phytother Res* **2017**, *31*, 555-567.
90. Sen, M.; Dash, B. A Review on Phytochemical and Pharmacological Aspects of *Cissus quadrangularis* L. *Int. J. Green Pharm.* **2012**, *6*, Art. No: 169. DOI: 10.4103/0973-8258.104924
91. Martinotti, S.; Ranzato, E. propolis: A New Frontier for Wound Healing? *Burns Trauma* **2015**, *3*, Art. No: 9. DOI: 10.1186/s41038-015-0010-z
92. Bolfa, P.; Vidrighinescu, R.; Petruta, A.; Dezmirean, D.; Stan, L.; Vlase, L.; Damian, G.; Catoi, C.; Filip, A.; Clichici, S. Photoprotective Effects of Romanian propolis on Skin of Mice Exposed to UVB Irradiation. *Food Chem. Toxicol.* **2013**, *62*, 329-342. DOI: 10.1016/j.fct.2013.08.078
93. Miryan, M.; Soleimani, D.; Dehghani, L.; Sohrabi, K.; Khorvash, F.; Bagherniya, M.; Sayedi, S.M.; Askari, G. The Effect of Propolis Supplementation on Clinical Symptoms in Patients with Coronavirus (COVID-19): A Structured Summary of a Study Protocol for a Randomised Controlled Trial. *Trials* **2020**, *21*, Art. No: 996. DOI: 10.1186/s13063-020-04934-7
94. Nazari-Bonab, H.; Jamilian, P.; Radkhah, N.; Zarezadeh, M.; Ebrahimi-Mameghani, M. The Effect of Propolis Supplementation in Improving Antioxidant Status: A Systematic Review and Meta-Analysis of Controlled Clinical Trials. *Phytother. Res.* **2023**, *37*, 3712-3723. DOI: 10.1002/ptr.7899
95. Hori, J.I.; Zamboni, D.S.; Carrão, D.B.; Goldman, G.H.; Berretta, A.A. The Inhibition of Inflammasome by Brazilian propolis (EPP-AF). *Evid. Based. Complement. Alternat. Med.* **2013**, *2013*, Art. No: 418508. DOI: 10.1155/2013/418508
96. Dalmonte, T.; Andreani, G.; Rudelli, C.; Isani, G. Efficacy of Extracts of Oleogum Resin of Boswellia in the Treatment of Knee Osteoarthritis: A Systematic Review and Meta-Analysis. *Phytother. Res.* **2024**, *38*, 5672-5689. DOI: 10.1002/ptr.8336
97. Abdel-Tawab, M.; Werz, O.; Schubert-Zsilavecz, M. *Boswellia Serrata*: An Overall Assessment of in Vitro, Preclinical, Pharmacokinetic and Clinical Data: An Overall Assessment of in Vitro, Preclinical, Pharmacokinetic and Clinical Data. *Clin. Pharmacokinet.* **2011**, *50*, 349-369.
98. Doaee, P.; Rajaei, Z.; Roghani, M.; Alaei, H.; Kamalinejad, M. Effects of *Boswellia Serrata* Resin Extract on Motor Dysfunction and Brain Oxidative Stress in an Experimental Model of Parkinson's Disease. *Avicenna J. Phytomed.* **2019**, *9*, 281-290.
99. Sailer, E.R.; Subramanian, L.R.; Rall, B.; Hoernlein, R.F.; Ammon, H.P.; Safayhi, H. Acetyl-11-Keto-Beta-Boswellic Acid (AKBA): Structure Requirements for Binding and 5-Lipoxygenase Inhibitory Activity. *Br. J.*

- Pharmacol. **1996**, 117, 615-618. DOI: 10.1111/j.1476-5381.1996.tb15235.x
100. Keppel Hesselink, J.M.; de Boer, T.; Witkamp, R.F. Palmitoylethanolamide: A Natural Body-Owned Anti-Inflammatory Agent, Effective and Safe against Influenza and Common Cold. *Int. J. Inflam.* **2013**, 2013, Art. No: 151028. DOI: 10.1155/2013/151028
101. Clayton, P.; Subah, S.; Venkatesh, R.; Hill, M.; Bogoda, N. Palmitoylethanolamide: A Potential Alternative to Cannabidiol. *J. Diet. Suppl.* **2023**, 20, 505-530. DOI: 10.1080/19390211.2021.2005733
102. Varrassi, G.; Rekatsina, M.; Leoni, M.L.G.; Cascella, M.; Finco, G.; Sardo, S.; Corno, C.; Tiso, D.; Schweiger, V.; Fornasari, D.M.M.; et al. A Decades-Long Journey of Palmitoylethanolamide (PEA) for Chronic Neuropathic Pain Management: A Comprehensive Narrative Review. *Pain Ther.* **2025**, 14, 81-101. DOI: 10.1007/s40122-024-00685-4
103. Petrosino, S.; Cordaro, M.; Verde, R.; Schiano Moriello, A.; Marcolongo, G.; Schievano, C.; Siracusa, R.; Piscitelli, F.; Peritore, A.F.; Crupi, R.; et al. Oral Ultramicronized Palmitoylethanolamide: Plasma and Tissue Levels and Spinal Anti-Hyperalgesic Effect. *Front. Pharmacol.* **2018**, 9, Art. No: 249. DOI: 10.3389/fphar.2018.00249.
104. Passavanti, M.B.; Alfieri, A.; Pace, M.C.; Pota, V.; Sansone, P.; Piccinno, G.; Barbarisi, M.; Aurilio, C.; Fiore, M. Clinical Applications of Palmitoylethanolamide in Pain Management: Protocol for a Scoping Review. *Syst. Rev.* **2019**, 8, Art. No: 9. DOI: 10.1186/s13643-018-0934-z
105. Artukoglu, B.B.; Beyer, C.; Zuloff-Shani, A.; Brenner, E.; Bloch, M.H. Efficacy of Palmitoylethanolamide for Pain: A Meta-Analysis. *Pain Physician* **2017**, 20, 353-362.
106. Nestmann, E.R. Safety of Micronized Palmitoylethanolamide (microPEA): Lack of Toxicity and Genotoxic Potential. *Food Sci. Nutr.* **2017**, 5, 292-309. DOI: 10.1002/fsn3.392
107. Schweiger, V.; Schievano, C.; Martini, A.; Polati, L.; Del Balzo, G.; Simari, S.; Milan, B.; Finco, G.; Varrassi, G.; Polati, E. Extended Treatment with Micron-Size Oral Palmitoylethanolamide (PEA) in Chronic Pain: A Systematic Review and Meta-Analysis. *Nutrients* **2024**, 16, Art. No: 1653. DOI: 10.3390/nu1611165.
108. Shamraiz, U.; Hussain, H.; Ur Rehman, N.; Al-Shidhani, S.; Saeed, A.; Khan, H.Y.; Khan, A.; Fischer, L.; Csuk, R.; Badshah, A.; et al. Synthesis of New Boswellic Acid Derivatives as Potential Antiproliferative Agents. *Nat. Prod. Res.* **2020**, 34, 1845-1852. DOI: 10.1080/14786419.2018.156429.
109. Sharma, S.; Gupta, S.; Khajuria, V.; Bhagat, A.; Ahmed, Z.; Shah, B.A. Analogues of Boswellic Acids as Inhibitors of Pro-Inflammatory Cytokines TNF- $\alpha$  and IL-6. *Bioorg. Med. Chem. Lett.* **2016**, 26, 695-698. DOI: 10.1016/j.bmcl.2015.11.03.
110. Mbiantcha, M.; Khalid, R.; Atsamo, D.A.; Njoku, I.S.; Mehreen, A.; Ateufack, G.; Hamza, D.; Nana, W.Y.; Naeem, R.U.; Izhar, A. Anti-Hypernociceptive Effects of Methanol Extract of Boswellia Dalzielii on STZ-Induced Diabetic Neuropathic Pain. *Adv. Tradit. Med.* **2020**, 20, 405-417. DOI: 10.1007/s13596-019-00411-y
111. Majeed, A.; Majeed, S.; Satish, G.; Manjunatha, R.; Rabbani, S.N.; Patil, N.V.P.; Mundkur, L. A Standardized Boswellia Serrata Extract Shows Improvements in Knee Osteoarthritis within Five Days—a Double-Blind, Randomized, Three-Arm, Parallel-Group, Multi-Center, Placebo-Controlled Trial. *Front. Pharmacol.* **2024**, 15, Art. No: 1428440. DOI: 10.3389/fphar.2024.1428440.
112. Roy, N.K.; Parama, D.; Banik, K.; Bordoloi, D.; Devi, A.K.; Thakur, K.K.; Padmavathi, G.; Shakibaei, M.; Fan, L.; Sethi, G.; et al. An Update on Pharmacological Potential of Boswellic Acids against Chronic Diseases. *Int. J. Mol. Sci.* **2019**, 20, Art. No: 4101. DOI: 10.3390/ijms20174101
113. Machado, J.L.; Assunção, A.K.M.; da Silva, M.C.P.; Dos Reis, A.S.; Costa, G.C.; Arruda, D. de S.; Rocha, B.A.; Vaz, M.M. de O.L.L.; Paes, A.M. de A.; Guerra, R.N.M.; et al. Brazilian Green propolis: Anti-Inflammatory Property by an Immunomodulatory Activity. *Evid. Based. Complement. Alternat. Med.* **2012**, 2012, Art. No: 157652. DOI: 10.1155/2012/157652
114. Kasote, D.; Bankova, V.; Viljoen, A.M. Propolis: Chemical Diversity and Challenges in Quality Control. *Phytochem. Rev.* **2022**, 21, 1887-1911. DOI: 10.1007/s11101-022-09816-1
115. Katiyar, D. Propolis: A natural biomaterial. *Mater. Today Proc.* **2023**, in press. DOI: 10.1016/j.matpr.2023.05.522
116. Xu, W.; Lu, H.; Yuan, Y.; Deng, Z.; Zheng, L.; Li, H. The Antioxidant and Anti-Inflammatory Effects of Flavonoids from Propolis via Nrf2 and NF- $\kappa$ B Pathways. *Foods* **2022**, 11, Art. No: 2439. DOI: 10.3390/foods11162439
117. Muthusami, S.; Ramachandran, I.; Krishnamoorthy, S.; Govindan, R.; Narasimhan, S. Cissus Quadrangularis Augments IGF System Components in Human Osteoblast like SaOS-2 Cells. *Growth Horm. IGF Res.* **2011**, 21, 343-348. DOI: 10.1016/j.ghir.2011.09.002.
118. Dhanasekaran, S. Phytochemical Characteristics of Aerial Part of Cissus Quadrangularis (L) and Its in-Vitro Inhibitory Activity against Leukemic Cells and Antioxidant Properties. *Saudi J. Biol. Sci.* **2020**, 27, 1302-1309. DOI: 10.1016/j.sjbs.2020.01.005
119. Guerra, J.M.; Hanes, M.A.; Rasa, C.; Loganathan, N.; Innis-Whitehouse, W.; Gutierrez, E.; Nair, S.; Banu, J. Modulation of Bone Turnover by Cissus Quadrangularis after Ovariectomy in Rats. *J. Bone Miner. Metab.* **2019**, 37, 780-795. DOI: 10.1007/s00774-018-0983-3
120. Tasadduq, R.; Gordon, J.; Al-Ghanim, K.A.; Lian, J.B.; Van Wijnen, A.J.; Stein, J.L.; Stein, G.S.; Shakoobi, A.R. Ethanol Extract of Cissus Quadrangularis Enhances Osteoblast Differentiation and Mineralization of Murine Pre-Osteoblastic MC3T3-E1 Cells: Effect of Herbal Extract on Osteoblast Differentiation. *J. Cell. Physiol.* **2017**, 232, 540-547. DOI: 10.1002/jcp.25449
121. Liao, L.; Zhu, W.; Tao, C.; Li, D.; Mao, M. Cissus Quadrangularis L Extract-Loaded Tricalcium Phosphate Reinforced Natural Polymer Composite for

- Guided Bone Regeneration. *J. Mater. Sci. Mater. Med.* **2023**, 34, Art. No: 33. DOI: 10.1007/s10856-023-06739-x
122. Sadeghnia, H.R.; Arjmand, F.; Ghorbani, A. Neuroprotective Effect of *Boswellia Serrata* and Its Active Constituent Acetyl 11-Keto-B-Boswellic Acid against Oxygen-Glucose-Serum Deprivation-Induced Cell Injury. *Acta Pol. Pharm.* **2017**, 74, 911-920.
123. Nakhaei, K.; Bagheri-Hosseini, S.; Sabbaghzade, N.; Behmadi, J.; Boozari, M. Boswellic Acid Nanoparticles: Promising Strategies for Increasing Therapeutic Effects. *Rev. Bras. Farmacogn.* **2023**, 33, 713-723. DOI: 10.1007/s43450-023-00405-7
124. Li, W.; Ren, L.; Zheng, X.; Liu, J.; Wang, J.; Ji, T.; Du, G. 3-O-Acetyl-11-Keto- B -Boswellic Acid Ameliorated Aberrant Metabolic Landscape and Inhibited Autophagy in Glioblastoma. *Acta Pharm. Sin. B.* **2020**, 10, 301-312. DOI: 10.1016/j.apsb.2019.12.012
125. Bartolucci, M.L.; Marini, I.; Bortolotti, F.; Impellizzeri, D.; Di Paola, R.; Bruschetta, G.; Crupi, R.; Portelli, M.; Militi, A.; Oteri, G.; et al. Micronized Palmitoylethanolamide Reduces Joint Pain and Glial Cell Activation. *Inflamm. Res.* **2018**, 67, 891-901. DOI: 10.1007/s00011-018-1179-y
126. Loi, S.; Pontis, E.; Cofelice, A.; Pirarba, V.; Fais, S.; Daniilidis, M.F. Effect of Ultramicrosized-Palmitoylethanolamide and Co-Microsized Palmitoylethanolamide/Polydatin on Chronic Pelvic Pain and Quality of Life in Endometriosis Patients: An Open-Label Pilot Study. *Int J Womens Health* **2019**, 11, 443-449.
127. Lama, A.; Pirozzi, C.; Severi, I.; Morgese, M.G.; Senzacqua, M.; Annunziata, C.; Comella, F.; Del Piano, F.; Schiavone, S.; Petrosino, S.; et al. Palmitoylethanolamide Dampens Neuroinflammation and Anxiety-like Behavior in Obese Mice. *Brain Behav. Immun.* **2022**, 102, 110-123. DOI: 10.1016/j.bbi.2022.02.008
128. Bueno-Silva, B.; Kawamoto, D.; Ando-Suguimoto, E.S.; Casarin, R.C.V.; Alencar, S.M.; Rosalen, P.L.; Mayer, M.P.A. Brazilian Red Propolis Effects on Peritoneal Macrophage Activity: Nitric Oxide, Cell Viability, pro-Inflammatory Cytokines and Gene Expression. *J. Ethnopharmacol.* **2017**, 207, 100-107. DOI: 10.1016/j.jep.2017.06.015
129. Šuran, J.; Cepanec, I.; Mašek, T.; Radić, B.; Radić, S.; Tlak Gajger, I.; Vlajnić, J. Propolis Extract and Its Bioactive Compounds-from Traditional to Modern Extraction Technologies. *Molecules* **2021**, 26, Art. No: 2930. DOI: 10.3390/molecules26102930
130. Javed, S.; Mangla, B.; Ahsan, W. From Propolis to Nanopropolis: An Exemplary Journey and a Paradigm Shift of a Resinous Substance Produced by Bees. *Phytother. Res.* **2022**, 36, 2016-2041. DOI: 10.1002/ptr.7435
131. Lv, L.; Cui, H.; Ma, Z.; Liu, X.; Yang, L. Recent Progresses in the Pharmacological Activities of Caffeic Acid Phenethyl Ester. *Naunyn. Schmiedeberg's Arch. Pharmacol.* **2021**, 394, 1327-1339. DOI: 10.1007/s00210-021-02054-w
132. Kumar, R.; Singh, S.; Saksena, A.K.; Pal, R.; Jaiswal, R.; Kumar, R. Effect of *Boswellia Serrata* Extract on Acute Inflammatory Parameters and Tumor Necrosis Factor- $\alpha$  in Complete Freund's Adjuvant-Induced Animal Model of Rheumatoid Arthritis. *Int. J. Appl. Basic Med. Res.* **2019**, 9, 100-106. DOI: 10.4103/ijabmr.IJABMR\_248\_18
133. Choi, Y.-J.; Jung, J.I.; Bae, J.; Lee, J.K.; Kim, E.J. Evaluating the Anti-Osteoarthritis Potential of Standardized *Boswellia Serrata* Gum Resin Extract in Alleviating Knee Joint Pathology and Inflammation in Osteoarthritis-Induced Models. *Int. J. Mol. Sci.* **2024**, 25, Art. No: 3218. DOI: 10.3390/ijms25063218
134. Majeed, M.; Nagabhushanam, K.; Lawrence, L.; Nallathambi, R.; Thiyagarajan, V.; Mundkur, L. *Boswellia Serrata* Extract Containing 30% 3-Acetyl-11-Keto-Boswellic Acid Attenuates Inflammatory Mediators and Preserves Extracellular Matrix in Collagen-Induced Arthritis. *Front. Physiol.* **2021**, 12, Art. No: 735247. DOI: 10.3389/fphys.2021.735247
135. Ammon, H.P.T. Boswellic Acids and Their Role in Chronic Inflammatory Diseases. *Adv. Exp. Med. Biol.* **2016**, 928, 291-327. DOI: 10.1007/978-3-319-41334-1\_13
136. De Carvalho Fm De, A.; Schneider, J.K.; De Jesus, C.; De Andrade, L.N.; Amaral, R.G.; David, J.M. Brazilian Red Propolis: Extracts Production, Physicochemical Characterization, and Cytotoxicity Profile for Antitumor Activity. *Biomolecules* **2020**, 10, Art. No: 726. DOI: 10.3390/biom10050726
137. Fonseca, L.; Ribeiro, M.; Schultz, J.; Borges, N.A.; Cardozo, L.; Leal, V.O.; Ribeiro-Alves, M.; Paiva, B.R.; Leite, P.E.C.; Sanz, C.L.; et al. Effects of Propolis Supplementation on Gut Microbiota and Uremic Toxin Profiles of Patients Undergoing Hemodialysis. *Toxins (Basel)* **2024**, 16, Art. No: 416. DOI: 10.3390/toxins16100416
138. Okamoto, Y.; Hara, T.; Ebato, T.; Fukui, T.; Masuzawa, T. Brazilian Propolis Ameliorates Trinitrobenzene Sulfonic Acid-Induced Colitis in Mice by Inhibiting Th1 Differentiation. *Int. Immunopharmacol.* **2013**, 16, 178-183. DOI: 10.1016/j.intimp.2013.04.004
139. Kurek-Górecka, A.; Keskin, Ş.; Bobis, O.; Felitti, R.; Górecki, M.; Otręba, M.; Stojko, J.; Olczyk, P.; Kolayli, S.; Rzepecka-Stojko, A. Comparison of the Antioxidant Activity of Propolis Samples from Different Geographical Regions. *Plants* **2022**, 11, Art. No: 1203. DOI: 10.3390/plants11091203
140. Cristiano, C.; Pirozzi, C.; Coretti, L.; Cavaliere, G.; Lama, A.; Russo, R.; Lembo, F.; Mollica, M.P.; Meli, R.; Calignano, A.; et al. Palmitoylethanolamide Counteracts Autistic-like Behaviours in BTBR T+tf/J Mice: Contribution of Central and Peripheral Mechanisms. *Brain Behav. Immun.* **2018**, 74, 166-175. DOI: 10.1016/j.bbi.2018.09.003
141. Roviezzo, F.; Rossi, A.; Caiazzo, E.; Orlando, P.; Riemma, M.A.; Iacono, V.M.; Guarino, A.; Ialenti, A.; Cicala, C.; Peritore, A.; et al. Palmitoylethanolamide Supplementation during Sensitization Prevents Airway Allergic Symptoms in the Mouse. *Front. Pharmacol.* **2017**, 8, Art. No: 857. DOI: 10.3389/fphar.2017.00857
142. Impellizzeri, D.; Di Paola, R.; Cordaro, M.; Gugliandolo, E.; Casili, G.; Morittu, V.M.; Britti, D.; Esposito, E.; Cuzzocrea, S. Adelmidrol, a Palmitoylethanolamide



- Analogue, as a New Pharmacological Treatment for the Management of Acute and Chronic Inflammation. *Biochem. Pharmacol.* **2016**, *119*, 27-41. DOI: 10.1016/j.bcp.2016.09.001
143. Pirozzi, C.; Coretti, L.; Opallo, N.; Bove, M.; Annunziata, C.; Comella, F.; Turco, L.; Lama, A.; Trabace, L.; Meli, R.; et al. Palmitoylethanolamide Counteracts High-Fat Diet-Induced Gut Dysfunction by Reprogramming Microbiota Composition and Affecting Tryptophan Metabolism. *Front. Nutr.* **2023**, *10*, Art. No: 1143004. DOI: 10.3389/fnut.2023.1143004
144. Wang, Z.; Singh, A.; Jones, G.; Aitken, D.; Laslett, L.L.; Hussain, S.; García-Molina, P.; Ding, C.; Antony, B. *Boswellia* for osteoarthritis. *Cochrane Database Syst Rev.* **2022**, 2022, Art. No: CD014969. DOI: 10.1002/14651858.CD014969
145. Ragab, E.A.; Abd El-Wahab, M.F.; Doghish, A.S.; Salama, R.M.; Eissa, N.; Darwish, S.F. The Journey of Boswellic Acids from Synthesis to Pharmacological Activities. *Naunyn. Schmiedeberg's Arch. Pharmacol.* **2024**, *397*, 1477-1504. DOI: 10.1007/s00210-023-02725-w
146. Shen, J.; Abu-Amer, Y.; O'Keefe, R.J.; McAlinden, A. Inflammation and Epigenetic Regulation in Osteoarthritis. *Connect. Tissue Res.* **2017**, *58*, 49-63. DOI: 10.1080/03008207.2016.1208655
147. Gong, Y.; Jiang, X.; Yang, S.; Huang, Y.; Hong, J.; Ma, Y.; Fang, X.; Fang, Y.; Wu, J. The Biological Activity of 3-O-Acetyl-11-Keto-B-Boswellic Acid in Nervous System Diseases. *Neuromolecular Med.* **2022**, *24*, 374-384. DOI: 10.1007/s12017-022-08707-0
148. Rajeshkumar, S.; Menon, S.; Kumar, V.; Ponnaniakamadeen, M.; Ali, D.; Arunachalam, K. Anti-Inflammatory and Antimicrobial Potential of *Cissus Quadrangularis*-Assisted Copper Oxide Nanoparticles. *J. Nanomater.* **2021**, 2021, Art. No: 5742981. DOI: 10.1155/2021/5742981
149. Coutinho de Almeida, R.; Ramos, Y.F.M.; Mahfouz, A.; den Hollander, W.; Lakenberg, N.; Houtman, E.; van Hoolwerff, M.; Suchiman, H.E.D.; Rodríguez Ruiz, A.; Slagboom, P.E.; et al. RNA Sequencing Data Integration Reveals an miRNA Interactome of Osteoarthritis Cartilage. *Ann. Rheum. Dis.* **2019**, *78*, 270-277. DOI: 10.1136/annrheumdis-2018-213882
150. Muthusami, S.; Senthilkumar, K.; Vignesh, C.; Ilangoan, R.; Stanley, J.; Selvamurugan, N.; Srinivasan, N. Effects of *Cissus Quadrangularis* on the Proliferation, Differentiation and Matrix Mineralization of Human Osteoblast like SaOS-2 Cells. *J. Cell. Biochem.* **2011**, *112*, 1035-1045. DOI: 10.1002/jcb.23016
151. Awari, V.S.; Barvkar, V.T.; Ade, A.B.; Borde, M.Y. Endophytic Fungi from *Cissus Quadrangularis* Plant a Promising Source of Bioactive Compounds. *Braz. J. Microbiol.* **2024**, *55*, 3733-3750. DOI: 10.1007/s42770-024-01500-0
152. Abdel-Tawab, M.; Werz, O.; Schubert-Zsilavecz, M. *Boswellia serrata*: an overall assessment of in vitro, preclinical, pharmacokinetic and clinical data. *Clin Pharmacokinet.* **2011**, *50*, 349-69. DOI: 10.2165/11586800-000000000-00000
153. Yu, G.; Xiang, W.; Zhang, T.; Zeng, L.; Yang, K.; Li, J. Effectiveness of *Boswellia* and *Boswellia* extract for osteoarthritis patients: a systematic review and meta-analysis. *BMC Complement. Med. Ther.* **2020**, *20*, Art. No: 225. DOI: 10.1186/s12906-020-02985-6
154. Pérez-Lozano, M.L.; Cesaro, A.; Mazor, M.; Esteve, E.; Berteina-Raboin, S.; Best, T.M.; Lespessailles, E.; Toumi, H. Emerging Natural-Product-Based Treatments for the Management of Osteoarthritis. *Antioxidants (Basel)* **2021**, *10*, Art. No: 265. DOI: 10.3390/antiox10020265
155. Kurek-Górecka, A.; Rzepecka-Stojko, A.; Górecki, M.; Stojko, J.; Sosada, M.; Swierczek-Zieba, G. Structure and Antioxidant Activity of Polyphenols Derived from Propolis. *Molecules* **2013**, *9*, 78-101. DOI: 10.3390/molecules19010078
156. Sforcin, J.M. Propolis and the Immune System: A Review. *J. Ethnopharmacol.* **2007**, *113*, 1-14. DOI: 10.1016/j.jep.2007.05.012
157. Mcalindon, T.E.; Lavalley, M.P.; Harvey, W.F. Effectiveness of Intra-Articular Corticosteroids for Knee Osteoarthritis: A Systematic Review. *Ann. Intern. Med.* **2017**, *166*, 255-266.
158. Gopukumar, K.; Raveendran, R.; Rao, M.N. Bone Healing Potential of *Cissus Quadrangularis*: A Systematic Review. *J. Ayurveda Integr. Med.* **2019**, *10*, 165-172.
159. Pande, S.; Pathak, P. Molecular Mechanisms of *Cissus Quadrangularis* in Bone Regeneration. *Phytother. Res.* **2016**, *30*, 1517-1526.
160. Coxib and traditional NSAID Trialists' (CNT) Collaboration; Bhala, N.; Emberson, J.; Merhi, A.; Abramson, S.; Arber, N.; Baron, J.A.; Bombardier, C.; Cannon, C.; Farkouh, M.E.; et al. Vascular and Upper Gastrointestinal Effects of Non-Steroidal Anti-Inflammatory Drugs: Meta-Analyses of Individual Participant Data from Randomised Trials. *Lancet* **2013**, *382*, 769-779. DOI: 10.1016/S0140-6736(13)60900-9
161. Hesselink, J.M.K.; Hekker, T.A. Therapeutic Utility of Palmitoylethanolamide in the Treatment of Neuropathic Pain Associated with Various Pathological Conditions: A Case Series. *J. Pain Res.* **2012**, *5*, 437-442. DOI: 10.2147/JPR.S32143
162. Paladini, A.; Fusco, M.; Cenacchi, T. Palmitoylethanolamide: A Potential Therapeutic Agent in Pain Management. *Clin. Drug Investig.* **2017**, *37*, 729-737.
163. Oo, W.M.; Yu, S.P.; Daniel, M.S.; Hunter, D.J. Disease-modifying drugs in osteoarthritis: current understanding and future therapeutics. *Expert Opin. Emerg. Drugs* **2018**, *23*, 331-347. DOI: 10.1080/14728214.2018.1547706
164. Ghouri, A.; Conaghan, P.G. Update on novel pharmacological therapies for osteoarthritis. *Ther. Adv. Musculoskelet. Dis.* **2019**, *11*, Art. No: 1759720x19864492. DOI: 10.1177/1759720X19864492
165. Kalayil, N.; Budar, A.A.; Dave, R.K. Nanofibers for Drug Delivery: Design and Fabrication Strategies. *BIO Integr.* **2024**, *5*, Art. No: 22. DOI: 10.15212/bioi-2024-0023